J.S. Department of the Interior J.S. Geological Survey

Field Tests of Polyethylene-Membrane
Diffusion Samplers for Characterizing
Volatile Organic Compounds in
Stream-Bottom Sediments, Nyanza
Chemical Waste Dump Superfund Site,
Ashland, Massachusetts

Vater-Resources Investigations Report 00-4108

Prepared in cooperation with the U.S. ENVIRONMENTAL PROTECTION AGENCY





Table 2. Concentrations of volatile organic compounds detected in vapor from vapor-diffusion samplers retrieved on February 16, 1999, near the Nyanza Chemical Waste Dump Superfund Site, Ashland, Massachusetts

[Volatile organic compound: Compound not detected at the reporting limit given in parentheses. PCE, tetrachloroethene; TCE, trichloroethene; trace, compound detected at concentration below reporting limit. Other compounds detected and comments: DCE, dichloroethene; EUPs, early unidentified peaks on gas chromatograph. dup, field duplicate sample; ppb v. parts per billion by volume; --, not analyzed]

Vapor		D-11-		V	olatile organic	compound	Other compared			
sampler name	Date installed	Bottom - material	Benzene TCE Toluene PCE Chlorobenzene (ppb v) (ppb v) (ppb v) (ppb v) (ppb v)				 Other compounds detected and comments 			
			m		Sudbury Riv	er in Millpon	nd			
V1	1-19-99	Cobbles and pebbles	(12)	18	(40)	(20)	(40)	EUPs		
V1z	1-19-99	Cobbles and pebbles	(12)	23	(40)	(20)	(40)	EUPs. Sampler placed 6 ft from V1		
V1a	1-19-99	Organic	(12)	37	(40)	(20)	(40)	Installed in ice-free water near bank		
V1a (dup)	1-19-99	Organic	(12)	49	(40)	(20)	(40)			
V1b	1-19-99	Organic	(12)	220	(40)	(20)	52	Installed in ice-free water near bank. EUPs, cis-DCE		
V2	1-19-99	Organic -very soft	(12)	560	(40)	(20)	960	At location of V2-C1 cluster.		
V2a	1-19-99	Organic	(12)	43	(40)	(20)	520	EUPs, cis-DCE		
V2a (dup)	1-19-99	Organic	(12)	80	(40)	(20)	490	EUPs, cis-DCE		
V2b	1-19-99	Organic	(12)	340	(40)	(20)	210	EUPs, cis-DCE		
V3	1-19-99	Organic	(12)	1,910	(40)	(20)	2,250	EUPs, cis-DCE		
V3a	1-19-99	Organic	(12)	23	(40)	(20)	1,270	EUPs, cis-DCE		
V3b	1-19-99	Organic	(12)	(12)	(40)	(20)	(40)	EUPs, cis-DCE		
V4	1-19-99	Organic	54	430	(40)	(20)	5,330	EUPs, cis-DCE		
V4b	1-19-99	Gravel	(12)	(12)	trace	(20)	(40)	EUPs, cis-DCE		
V5	1-19-99	Gravel	(12)	(12)	(40)	(20)	(40)	EUPs		
				Sudb	oury River Do	wnstream fro	m Dam			
V6	1-19-99	Gravel	(12)	(12)	(40)	(20)	(40)	EUPs		
V7-2	1-19-99	Gravel	(12)	(12)	(40)	(20)	78	EUPs		
V7a-2	1-19-99	Gravel	(12)	(12)	(40)	(20)	(40)	EUPs		
V7a-2 (dup)	1-19-99	Gravel	(12)	(12)	(40)	(20)	(40)	EUPs		
V8-2	1-19-99	Gravel	(12)	(12)	(40)	(20)	(40)	EUPs		
					Former M	Iill Raceway				
V7-1	1-19-99	Organic	18	590	(40)	(20)	940	EUPs, cis-DCE. Placed at location of V7-C2 cluster.		
V7a-1	1-19-99	Organic	55	35	(40)	(20)	1,780	EUPs, cis-DCE		
V7a-1 (dup)	1-19-99	Organic	51	37	(40)	(20)	1,630	EUPs, cis-DCE		
V7b-1 (lost)	1-19-99	Organic			·		, 	Placed between PW7-1 and PW8-1 in raceway		
V8-1	1-19-99	Gravel	15	34	(40)	(20)	130	EUPs, cis-DCE		
V8a-1	1-19-99	Gravel	15	26	(40)	(20)	93	EUPs, cis-DCE. Located about 1 ft from V8-1.		

Although data are limited, results from vapor samplers indicate that the VOCs in bottom sediments along the Sudbury River were present in low concentrations (chlorobenzene detected at a concentration of 78 µg/L at location V7-2) or absent downstream from the dam at the time of the study. The presence of elevated concentrations in samplers placed along the raceway (table 2, fig. 2), however, is consistent with the mapped extent of the contaminant plume in ground water (fig. 2). VOCs detected at location 08-01 in the raceway beyond the mapped extent of the plume may reflect the presence of VOCs in surface water at this location (Roy F. Weston, Inc., 1999b) and an exchange of VOCs between surface water and ground water. Similar conditions for the Royal River near Gray, Maine, downstream from a TCE plume in ground water, have been described by Lyford and others (1999).

EQUILIBRATION TIME FOR VOCS IN DIFFUSION SAMPLERS

Results of analyses for vapor samplers retrieved about once a week from the 4 clusters were not conclusive with regard to equilibration time (table 3, figs. 5A and 5B). Results of experiments elsewhere have shown that concentrations of VOCs in vapordiffusion samplers equilibrate within less that 24 hours when placed in a solution that has a known concentration of VOCs. Equilibration after emplacement can be within a period of 24 hours when placed in sandy materials, but can take several days when placed in fine-grained materials (D.A. Vroblesky, U.S. Geological Survey, written commun., 1999). Apparent general increases in concentrations of chlorobenzene and TCE with time at clusters V2-C1 and V7-C2 (table 3, fig. 5) may reflect slow equilibration times of 3 weeks or more at these

Table 3. Concentrations of volatile organic compounds detected in clusters of vapor-diffusion samplers retrieved during January and February 1999 near the Nyanza Chemical Waste Dump Superfund Site, Ashland, Massachusetts

[Volatile organic compound: Compound not detected at the reporting limit given in parentheses. PCE, tetrachloroethene; TCE, trichloroethene; trace, compound detected at concentration below reporting limit. Other compounds detected and comments: DCE, dichloroethene; EUPs, early unidentified peaks on gas chromatograph. ppb v, parts per billion by volume]

Vapor sampler name	Date installed	Date retrieved			Volatile	organic co			
			Bottom material	Benzene (ppb v)	TCE (ppb v)	Toluene (ppb v)	PCE (ppb v)	Chloro- benzene (ppb v)	Other compounds detected and comments
V2-C1a	1-20-99	1-28-99	Organic	(20)	380	(50)	(30)	650	EUPs, cis-DCE
V2-C1b	1-19-99	2- 2-99	Organic	(12)	610	(40)	(20)	520	EUPs, cis-DCE
V2-C1c	1-19-99	2- 8-99	Organic	(12)	990	(40)	(20)	940	EUPs, cis-DCE; other unidentified peaks
V2	1-19-99	2-16-99	Organic - very soft	(12)	560	(40)	(20)	960	At location of V2-C1 cluster.
V2-C2a	1-19-99	1-28-99	Organic	(12)	330	(50)	(30	160	EUPs, cis-DCE
V2-C2b	1-19-99	2- 2-99	Organic	(12)	770	(50)	(30)	780	EUPs, cis-DCE
V2-C2c	1-19-99	2- 8-99	Organic	(12)	690	(40)	(20)	520	EUPs, cis-DCE; other unidentified peaks
V7-C1a	1-19-99	1-28-99	Organic	(20)	490	(50)	(30)	720	EUPs, cis-DCE
V7-C1b	1-19-99	2- 2-99	Organic	12	73	(40)	(20)	610	EUPs, cis-DCE
V7-C1c	1-19-99	2- 8-99	Organic	trace	170	(40)	(20)	500	EUPs, cis-DCE
V7-C2a	1-20-99	1-28-99	Organic	(20)	150	(50	(30)	52	EUPs, cis-DCE
V7-C2b	1-19-99	2- 2-99	Organic	(12)	390	(40)	(20)	460	EUPs, cis-DCE
V7-C2c	1-19-99	2-8-99	Organic	(12)	1,330	(40)	40	920	EUPs, cis-DCE
V7-1	1-19-99	2-16-99	Organic	18	590	(40)	(20)	940	EUPs, cis-DCE. Placed at location of V7-C2 cluster.

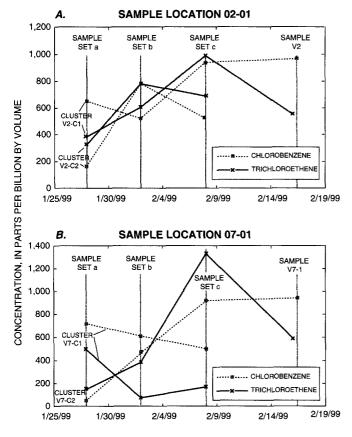


Figure 5. Concentrations of chlorobenzene and trichloroethene detected in vapor-diffusion samplers at (A) sample location 02-01 and (B) sample location 07-01, January and February 1999, Ashland, Massachusetts.

locations. Other factors, however, that may cause apparent changes in concentrations with time include varying river stages (fig. 3) that affect rates of groundwater flow to or from the stream, spatial variations over short distances, or a combination of these factors. Variability can also be attributed, in part, to the sampling and analytical methods as indicated by differences observed between duplicate samples at sampler V2a (table 2) where chlorobenzene concentrations were similar in the two samples but TCE concentrations differed by a factor of 2. Differing characteristics of bottom sediments (table 2) had no obvious effects on equilibration time. No VOCs were detected in trip blanks transported to the laboratory with cluster samples.

The vapor- and water-diffusion pairs at the locations of clusters V2-C1 and V7-C2 (fig. 4) that were retrieved on February 16, 1999, one week after retrieving the last sample in each cluster of 3 vapor samples at the same locations, provide a fourth sample for comparison to samples collected previously. The

concentration of chlorobenzene in the vapor sampler of each pair was about the same as the concentration in the last sampler retrieved, but the concentration of TCE decreased by about one half (table 3). The presence of the water-diffusion sampler may have affected concentrations of VOCs in the vapor-diffusion samplers for unknown reasons. Of possible significance was the observation that several vapordiffusion samplers were iron-stained upon retrieval, indicating that oxygen was diffusing from air in the bottles to water outside the bottles and altering the geochemistry of the solution locally. It is not known, however, if this process affected concentrations of VOCs in air relative to their concentrations in water.

The possibility that pore water chemistry, including concentration of VOCs, was changing at cluster locations during the study period is supported by specific-conductance measurements for water from seepage meters on January 18 and again on February 15-16, 1999. For example, the specific conductance increased from 360 to 450 µS/cm at seepage meter PW02-01 and from 910 to 1,020 µS/cm at seepage meter PW07-01 between measurements. By contrast, the specific conductance of water in the Sudbury River decreased from about 320 to 250 µS/cm between measurements (Roy F. Weston, Inc., 1999b, Appendix B).

COMPARISON OF CHEMICAL **ANALYTICAL RESULTS**

VOCs were detected in bottom sediments along the Sudbury River and mill raceway using vapor diffusion-samplers discussed above, water-diffusion samplers, seepage meters, and chemical analyses of the sediments. Each sampling method has its attributes in terms of ease, expense, and reliability of results. This section will qualitatively compare results from the four methods using tables and graphs. Concentrations of VOCs for samples collected by the four methods at each location are given in table 4.

Vapor- and Water-Diffusion Samples

When in equilibrium, the concentration of VOCs in air should correlate with the concentrations in water, as predicted by Henry's Law. Factors that will affect the relation between concentrations in air and concen-

Table 4. Concentrations of volatile organic compounds in vapor-diffusion samplers, water-diffusion samplers, seepage meters, and sediments, December 1998 to February 1999, Nyanza Chemical Waste Dump Superfund Site, Ashland, Massachusetts

[Data for seepage meters and sediment from R.F. Weston, Inc., 1999. Sample type and units: PW, seepage meter sample; SD, sediment sample; V, vapor-diffusion sample; W, water-diffusion sample. V and W samples collected on February 16, 1999; PW samples collected on February 15–16, 1999; SD samples collected on December 26–30, 1998. Compound not detected at the reporting limit given in parentheses. J, the compound was detected but near the practical quantitation limit; ns, not sampled or not analyzed; ppb v, parts per billion by volume; µg/kg, micrograms per kilogram; µg/L, micrograms per liter]

Volatile organic	Sample type and units	Sample location (fig. 1)										
compound		01–01	02–01	03–01	04–01	05–01	06–01	0701	07–02	08–01	08-02	
Acetone	V (ppb v)	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	
	W (μg/L)	4 J	(5)	5	4J	6	5 J	4 J	3J	6	ns	
	PW (µg/L)	4J	(5)	3 J	3Ј	(5)	(5)	(5)	(5)	(5)	(5.0)	
	SD (µg/kg)	910 J	1,000	220J	210J	520	260	340	(240)	(220)	(700)	
Benzene	V (ppb v)	(10)	(10)	(10)	54	(10)	(10)	18	(10)	15	(10)	
	W (μg/L)	(1.0)	.8J	1	4	(1.0)	(1.0)	1	(1.0)	.8Ј	ns	
	PW (μg/L)	(1.0)	.6J	.8J	3	(1.0)	(1.0)	2	(1.0)	(1.0)	(1.0)	
	SD (µg/kg)	(1,600)	(1.000)	(310)	(300)	(410)	(240)	(340)	(240)	(220)	(700)	
2-Butanone	V (ppb v)	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	
	W (μg/L)	4 J	(5)	5	(5)	(5)	(5)	(5)	(5)	(5)	ns	
	PW (µg/L)	(5)	(5)	(5)	(5)	(5)	(5)	(5)	(5)	(5)	(5)	
	SD (µg/kg)	(1,600)	(1,000)	260J	200J	(410)	(240)	480	(240)	(220)	(700)	
Chlorobenzene	V (ppb v)	(40)	960	2,250	5,330	(40)	(40)	940	78	130	(40)	
	W (μg/L)	(1.0)	50	120	180	(1.0)	(1.0)	61	.8J	5	ns	
	PW (µg/L)	(1.0)	38	74	140	(1.0)	.9Ј	58	(1.0)	2	(1.0)	
	SD (µg/kg)	(310)	1,300	120	360	(82)	(49)	980	(49)	(45)	140)	
Chloroform	V (ppb v)	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	
	$W(\mu g/L)$	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	ns	
	PW (µg/L)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	.6J	(1.0)	(1.0)	(1.0)	(1.0)	
	SD (µg/kg)	(310)	(210)	(62)	(61)	(82)	(49)	(69)	(49)	(45)	(140)	
Chloromethane	V (ppb v)	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	
	W (μg/L)	(2)	(2)	(2)	(2)	(2)	(2)	(2)	(2)	(2)	ns	
	PW (µg/L)	(2)	(2)	4	(2)	(2)	9	(2)	(2)	(2)	4	
	SD (µg/kg)	(620)	(420)	(120)	(120)	(160)	(98)	(140)	(98)	(90)	(280)	
1,1-Dichloroethene	V (ppb v)	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	
	W (μg/L)	(1.0)	(1.0)	1	1	(1.0)	(1.0)	2	(1.0)	(1.0)	ns	
	PW (µg/L)	(1.0)	.6J	.5J	1	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	
	SD (µg/kg)	(310)	(210)	(62)	(61)	(82)	(49)	(69)	(49)	(45)	(140)	

Table 4. Concentrations of VOCs in vapor-diffusion samplers, water-diffusion samplers, seepage meters, and sediments, December 1998 to February 1999, Nyanza Chemical Waste Dump Superfund site, Ashland, Massachusetts—*Continued*

Volatile organic	Sample type	Sample location (fig. 1)										
compound	and units	01–01	02–01	0301	04–01	05–01	06–01	07–01	07–02	08-01	08–02	
cis-1,2-Dichloroethene	V (ppb v)	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	
(cis-DCE) (total 1,2-	W (µg/L)	(2.0)	25	70	70	(2.0)	(2.0)	62	(2.0)	4	ns	
Dichloroethene for	PW (µg/L)	(2.0)	47	44	60	(2.0)	(2.0)	67	(2.0)	5	(2.0)	
sediment samples)	SD (µg/kg)	(310)	230	(72)	44J	(82)	(49)	240	(49)	(45)	(140)	
trans-1,2-Dichloroethene	V (ppb v)	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	
	W (μg/L)	(2.0)	(2.0)	1J	(2.0)	(2.0)	(2.0)	1 J	(2.0)	(2.0)	ns	
	PW (µg/L)	(2.0)	(2.0)	(2.0)	(2.0)	(2.0)	(2.0)	1J	(2.0)	(2.0)	(2.0)	
	SD (µg/kg)	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	
Ethylbenzene	V (ppb v)	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	
	W (μg/L)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	4	(1.0)	ns	
	PW (µg/L)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)		(1.0)	
	SD (µg/kg)	(1,600)	(1,000)	(310)	(300)	(410)	(240)	(340)	(240)	(220)	(700)	
Tetrachloroethene (PCE)	V (ppb v)	(20)	(20)	(20)	(20)	(20)	(20)	(20)	(20)	(20)	(20)	
	W (µg/L)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	.5J	(1.0)	(1.0)	ns	
	PW (µg/L)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	
	SD (µg/kg)	(310)	(1,000)	(62)	(61)	(82)	(49)	(69)	(49)	(45)	(140)	
Toluene	V (ppb v)	(40)	(40)	(40)	(40)	(40)	(40)	(40)	(40)	(40)	(40)	
	W (μg/L)	(1.0)	(1.0)	.7Ј	.8J	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	ns	
	PW (µg/L)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	
	SD (µg/kg)	(1,600)	(1,000)	(310)	(300)	(410)	(240)	(340)	(240)	(220)	(700)	
Trichloroethene (TCE)	V (ppb v)	18	560	1,910	430	(12)	(12)	590	(12)	34	(12)	
	W (μg/L)	1	5	110	11	(1.0)	(1.0)	22	(1.0)	4	ns	
	PW (µg/L)	1	16	60	16	(1.0)	(1.0)	35	(1.0)	5	(1.0)	
	SD (µg/kg)	(310)	240	(62)	(61)	(82)	(49)	700	(49)	(45)	(140)	
Vinyl chloride	V (ppb v)	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	
	W (μg/L)	(2.0)	13	9	67	(2.0)	(2.0)	19	(2.0)	2	ns	
	PW (µg/L)	(2.0)	6	7	58	(2.0)	(2.0)	23	(2.0)	3	(2.0)	
	SD (µg/kg)	(620)	(420)	(120)	(120)	(160)	(98)	(140)	(98)	(90)	(280)	
Total Xylenes	V (ppb v)	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	
	W (μg/L)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	29	(1.0)	ns	
	PW (µg/L)	(1.0)	0.7J	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	(1.0)	
	SD (µg/kg)	(1,600)	(1,000)	(310)	(300)	(410)	(240)	(340)	(240)	(220)	(700)	

trations in water are temperature, pressure, and molecular weight of a compound. Calculations made using information provided by Rathbun (1998) about Henry's Law constants and their variation with temperature indicate that TCE concentrations in air, in units of parts per billion by volume, can be 20 to 30 times its concentration in water, in micrograms per liter, for a temperature range of 1°C to 10°C. For chlorobenzene, the concentration in air can be from 10 to 16 times its concentration in water for that same temperature range.

Several VOCs were detected in water from water-diffusion samplers, but the principal compounds and maximum concentrations detected were for chlorobenzene (180 µg/L at location 04-01), TCE (110 µg/L at location 03-01), cis-DCE (70 µg/L at locations 03-01 and 04-01), and vinyl chloride (67 µg/L at location 04-01) (table 4). Chemical analyses for water included a larger number of chemicals than the 5 target compounds for vapordiffusion samples. Table 4 reports all of the VOCs that were detected in the water-diffusion samples. In general, the target compounds detected in vapor samples were consistent with compounds detected in water samples at each sampling location. Exceptions were benzene, which was detected in vapor samples at only three of the five locations where it was detected in water samples, and toluene, which was detected at low concentrations (less than 1 µg/L) at two locations (03-01 and 04-01) in water samples but not in vapor samples at the same locations.

Figure 6 shows the relation of concentrations in vapor to concentrations in water at vapor- and waterdiffusion sampler pairs for chlorobenzene and TCE, the major compounds detected in vapor and water. Also shown is the range of concentrations that might be expected on the basis of Henry's Law, assuming a temperature range of 1°C to 10°C and samples collected at atmospheric pressure. The temperature range from 1°C to 10°C represents a possible range during the study period from near 0°C at times in surface water to a maximum of 10°C in ground water. Although a fair correlation is apparent for the two types of samples, deviations from concentrations predicted by Henry's Law indicate that either the vapor samples, water samples, or both, had not equilibrated with sediment pore water. Uncertainties in chemical analytical results could also affect the apparent relation between concentrations in air and concentrations in water.

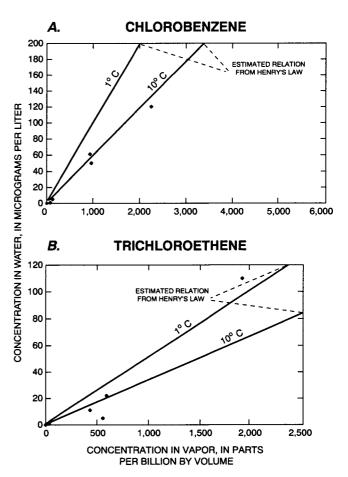


Figure 6. Concentrations of (A) chlorobenzene and (B) trichloroethene in vapor- and water-diffusion samplers, February 16, 1999, Ashland, Massachusetts.

Water-Diffusion Samples and Seepage-**Meter Samples**

Results of chemical analyses of water samples extracted from seepage meters (Roy F. Weston, Inc., 1999b) are included in table 4 for comparison to results from water-diffusion samplers. In general, the principal compounds detected by the two methods are consistent, and concentrations for the major constituents are similar, as shown in figure 7. At concentrations of individual constituents greater than about 50 µg/L, the concentrations in water-diffusion samples were generally higher than concentrations in seepage-meter samples. This indicates that analyses of water from seepage meters may have underestimated concentrations in pore water. If the water-diffusion samplers had not fully equilibrated, as discussed above, then both methods may underestimate actual concentrations in pore water, at least at the higher concentrations.

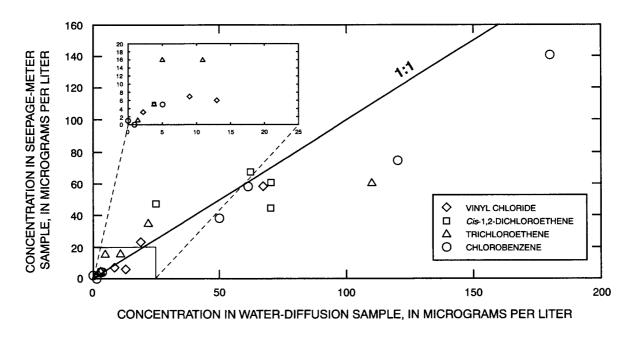


Figure 7. Concentrations of selected volatile organic compounds detected in water-diffusion samplers and concentrations in water from seepage meters, Ashland, Massachusetts.

Several compounds that were present near the detection limit in some diffusion samplers were not detected in nearby seepage meters, and some compounds were detected in seepage meters but not in diffusion samplers. Compounds that were present in concentrations appreciably above the detection limit were total xylenes, which were detected in diffusion sampler W7-2 but not in the nearby seepage meter, and chloromethane, detected in water from seepage meters PW03-01 and PW06-01 but not in nearby diffusion samplers. No patterns that might be related to the sampling method were apparent. Differences in compounds detected and concentrations may be attributable, in part, to variations in plume chemistry over distances of several feet between the seepage meters and diffusion samplers.

Water-Diffusion Samples and Sediment Samples

Results of chemical analyses of sediment samples collected at the locations of seepage meters (Roy F. Weston, Inc., 1999b) are included in table 4 for comparison to results from other sampling methods. In general, the principal VOCs detected in sediment are consistent with those detected in water-diffusion samplers (and water from seepage meters as discussed

above). Examination of concentrations reported in table 4 [accounting for different units for the two types of samples (µg/L for water and µg/kg for sediment)], however, indicate a poor correlation for the two methods. At several locations, such as 01-01, 03-01, 04-01, and 08-01, TCE was present in water-diffusion samples but was not detected in sediment samples. Other compounds that were detected in water-diffusion samples but not in sediment included vinyl chloride, 1,1,-dichloroethene, and benzene. The apparent absence of some compounds in sediments may result from relatively high quantitation limits for the sediment analyses and a shallower sampling depth (6 in. or less) than the depths of the water-diffusion samplers (6 to 8 in.). These results indicate that the other three sampling methods yielded a better representation of the VOCs present in sediments than chemical analyses of the sediments. Concentrations of total organic carbon at the sampling locations considered here ranged from 4,900 to 116,000 mg/kg (Roy F. Weston, Inc., 1999b). The relative concentration of VOCs in water samples and sediment samples are not obviously affected by the concentration of total organic carbon in sediment, although this possibility cannot be thoroughly assessed with the limited data available.

USE OF DIFFUSION SAMPLERS AS RECONNAISSANCE TOOLS

This study has reaffirmed that vapor-diffusion samplers placed in bottom sediments are useful reconnaissance tools for determining the distribution of VOCs in ground water and variations in concentrations across the width of a contaminant plume that discharges to a surface-water body. Others who have demonstrated uses of vapor-diffusion samplers for this purpose include Vroblesky and others (1996), Savoie and others (1999), and Lyford and others (1999). Sampling results can also provide useful information about ground-water flow patterns. For example, the vapor samplers used for this study, although few in number, confirmed a likely losing reach in the downstream end of the mill pond and indicated minimal discharge of contaminants to the Sudbury River downstream from the dam. This scenario differs from a previous conceptual model of contaminant movement that showed the plume of contaminants extending to the Sudbury River (fig. 2). Although beyond the scope of this study, a network of vapordiffusion samplers placed within the mill pond area might have identified discharge points for ground water that contained higher concentrations of VOCs, consistent with concentrations in deep ground water, than those apparent near the shore. Networks of vapordiffusion samplers have been used to delineate groundwater discharge areas and flow patterns near ponds on Cape Cod (D.R. LeBlanc, U.S. Geological Survey, oral commun., 1999). Diffusion samplers may have limitations where VOCs are present in surface water, because the source of VOCs could be either surface water or ground water.

Experience at the Nyanza site and other sites in New England has demonstrated that one person can construct as many as 100 samplers per day, and a crew of 3 can install 50 to 150 samplers per day; these totals depend somewhat on ease of access to the study area and character of the bottom sediments. Retrieval times and personnel requirements are less for retrieval than for installation. An analyst using a gas chromatograph can analyze about 50 vapor samples in a day. Therefore, a considerable amount of information can be generated in a short time relative to other sampling methods.

Comparison of VOCs in vapor-diffusion samplers to VOCs in adjacent water-diffusion samplers indicates that different equilibration times after

emplacement might limit the use of either method for reliably determining actual concentrations of VOCs in pore water. At present, the equilibration times for various types of bottom materials and various hydrologic conditions are not well defined. Other factors that might affect concentrations and interpretation of results are time-varying hydrologic conditions that affect the flux of contaminated ground water to streams, variations in concentrations over short distances, and exchanges between ground water and surface water in the hyporheic zone, which is the subsurface zone where stream water flows through short segments of its adjacent beds and banks (Winter and others, 1998).

The results of this study indicates that waterdiffusion samplers are a viable alternative to seepage meters for identifying the types of VOCS that are present in bottom sediments. A water-diffusion sampler can be constructed, installed, and retrieved in a total time of an hour or less. This contrasts with a time of several hours for installation and sampling of seepage meters. Seepage meters or other techniques for sampling pore water, however, are needed to determine concentrations of other contaminants such as metals or SVOCs in bottom sediments.

Elevated concentrations of SVOCs and metals in ground water near the Nyanza site are associated with elevated concentrations of VOCs (Roy F. Weston, Inc., 1998). This condition is commonly observed at contaminated sites. In areas where VOCs are indicators of other contaminants in ground water, vapor-diffusion samplers can quickly and economically identify "hot spots" and guide the sampling of bottom sediments for other contaminants.

Vapor- and water-diffusion samplers may be useful for assessing geochemical processes such as biodegradation in bottom sediments. For example, results from water-diffusion samplers and seepage meters identified elevated concentrations of vinyl chloride (9 to 67 µg/L in water-diffusion samples) at four locations. Concentrations of vinyl chloride in water from wells near the river and raceway where ground water is discharging to surface water, however, are generally less than 2 µg/L. This concentration pattern indicates possible transformation of TCE by biodegradation as ground water moves upward through the organic-rich bottom sediments toward surface water. Biodegradation of chlorinated VOCs as ground water flows vertically upward through wetland sediment has been described by Lorah and Olsen (1999). A network of vapor- or water-diffusion

samplers could be useful for identifying active areas of biodegradation for chlorinated VOCs. Networks of vapor-diffusion samplers may also be useful for studying patterns of exchanges between surface water and ground water in the hyporheic zone, where VOCs are known to be discharging from ground water to surface water.

Because results from this study and studies in other New England settings have confirmed the value of vapor-diffusion samplers as simple and economical reconnaissance tools, further testing for this purpose is not warranted. If a goal is to determine actual concentrations of VOCs at the interface between ground water and surface water, however, additional studies are needed to evaluate equilibration times in various geohydrologic settings. Also needed to accomplish this goal is a better understanding of variations in concentrations over short distances of a few feet and possible changes in concentrations with changing hydrologic conditions. Networks of closelyspaced vapor samplers retrieved and analyzed over an extended time in several geohydrologic settings could provide useful insights on spatial and temporal variability.

SUMMARY

Polyethylene-membrane water-to-vapor (vapor) and water-to-water (water) diffusion samplers were installed during a pilot study in January 1999 in bottom sediments along the Sudbury River and former mill raceway to determine if vapor samplers would be useful in this setting for delineating a plume of contaminants in ground water near the river and raceway, to evaluate equilibration time for vapordiffusion samplers, and to determine if diffusion samplers might be an alternative to seepage meters (inverted steel drums) and sediment sampling for evaluating concentrations of VOCs in bottom sediments.

VOCs were detected in all but 5 of 21 vapordiffusion samplers (excluding samples from clusters and duplicate samples). Of 5 tested compoundsbenzene, trichloroethene, toluene, tetrachloroethene, and chlorobenzene—chlorobenzene and trichloroethene were detected most frequently. The distribution of VOCs was generally consistent with the mapped plume of contaminants in ground water. The

absence of VOCs in the downstream part of the mill pond was consistent with water-level well data, which indicated that water was leaking from the pond to the surficial aquifer in this area. The general absence of VOCs along the Sudbury River downstream from the dam indicated that the discharge of VOCs to the river was limited at the time of the study. Result from the experiment to evaluate equilibration times for vapordiffusion samplers were inconclusive because of changing hydrologic conditions that may have affected concentrations of VOCs, possible variations in concentrations of VOCs over short distances, and imprecise sampling and analytical methods. The limited data from this study, however, indicated that equilibration times may exceed 3 weeks in some settings.

Analyses of water from water-diffusion samplers included a larger suite of chemicals than analyses for vapor from vapor-diffusion samplers. The principal compounds and maximum concentrations detected in water-diffusion samplers were chlorobenzene (180 µg/L), trichloroethene (110 µg/L), cis 1,2dichloroethene (70 µg/L), and vinyl chloride (67 µg/L). Highest concentrations of TCE and chlorobenzene in water samples were found at the same locations as the highest concentrations detected in vapor samples. Concentrations of chlorobenzene and TCE detected in vapor-diffusion samplers generally correlated with concentrations detected in water-diffusion samplers. Also, the VOCs and concentrations detected in waterdiffusion samplers were generally consistent with those detected in water from seepage meters.

Results from this pilot study reaffirm results from previous studies that diffusion samplers are a simple and economical means of identifying "hot spots" for contaminants in bottom sediments, and can provide insights on pathways for contaminants near surface-water bodies. Diffusion samplers may also be useful for studying variations in concentrations of VOCs across short distances, variations with time and changing hydrologic conditions, and processes such as chemical transformations and exchanges between surface water and ground water in the hyporheic zone.

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